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# Oxidative Stress and Antioxidant Therapy in Critically Ill Polytrauma Patients with Severe Head Injury

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# ABSTRACT

Traumatic Brain Injury (TBI) is one of the leading causes of death among critically ill patients from the Intensive Care Units (ICU). After primary traumatic injuries, secondary complications occur, which are responsible for the progressive degradation of the clinical status in this type of patients. These include severe inflammation, biochemical and physiological imbalances and disruption of the cellular functionality. The redox cellular potential is determined by the oxidant/antioxidant ratio. Redox potential is disturbed in case of TBI leading to oxidative stress (OS). A series of agression factors that accumulate after primary traumatic injuries lead to secondary lesions represented by brain ischemia and hypoxia, inflammatory and metabolic factors, coagulopathy, microvascular damage, neurotransmitter accumulation, blood-brain barrier disruption, excitotoxic damage, blood-spinal cord barrier damage, and mitochondrial dysfunctions. A cascade of pathophysiological events lead to accelerated production of free radicals (FR) that further sustain the OS. To minimize the OS and restore normal oxidant/antioxidant ratio, a series of antioxidant substances is recommended to be administrated (vitamin C, vitamin E, resveratrol, N-acetylcysteine). In this paper we present the biochemical and pathophysiological mechanism of action of FR in patients with TBI and the antioxidant therapy available.

Keywords: antioxidant therapy, oxidative stress, traumatic brain injury, multiple trauma patients

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# **INTRODUCTION**

A high percentage of poly-trauma patients present with severe central nervous system (CNS) trauma. Traumatic Brain Injury (TBI) and spinal cord injury (SCI) are responsible for increased mortality and morbidity in most trauma patients. Primary brain injury is characterized by contusions caused by direct impact and include haematomas, intra-parenchymal contusions and bleeding from vascular damage [1]. Primary injuries and pathophysiological imbalances lead to a series of aggressive factors that produce secondary injuries. These include brain ischemia and hypoxia, inflammatory factors, metabolic factors, coagulopathy, microvascular damage, neurotransmitter accumulation, blood-brain barrier disruption, excitotoxic lesions, blood-spinal cord barrier damage, and mitochondrial dysfunction [2,3]. A number of specific aspects of the critically ill patient is associated with TBI, such as mechanical ventilation, suppression of the immune response and infections, and these can lead to a significant increase in the rate of morbidity and mortality. Also, in the case of the critically ill patient, TBI can significantly affect the clinical progress due to the generated changes in systemic conditions such as acute respiratory distress syndrome (ARDS), Takotsubo cardiomyopathy and consumptive coagulopathy. Also, recent studies indicate that Multiple organ dysfunction syndrome (MODS) can occur after severe TBI [1-4].

The consequence of both primary and secondary lesions is represented by increased synthesis of free

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radicals (FR) that induce oxidative stress (OS) [4]. FR aggressive action lead to physiological imbalances of the central nervous system, significantly reducing the survival rate of critical trauma patients.

In this paper we present the biochemical and pathophysiological implications of OS in patients with severe TBI. Moreover, a summary of the existing antioxidant therapy aimed at improving the therapeutical management of critically ill trauma patients, is given.

# BIOCHEMICAL ASPECTS OF OXIDATIVE STRESS IN CASE OF TRAUMATIC BRAIN INJURY

FR are extremely reactive chemical species. Increased FR biosynthesis capacity, and its reactivity towards macromolecules such as lipids, proteins and nucleic acids, have an aggressive impact for the human body (Figure 1).

FR produced in TBI are reactive species of oxygen and nitrogen [5]. Reactive oxygen species (ROS) include superoxide anions ( $O_2^-$ ), hydroxyl radical (HO<sup>-</sup>), peroxyl (RO<sub>2</sub><sup>-</sup>), hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>), hypochlorous acid (HOCl). Reactive nitrogen species (RNS) are represented by peroxynitrite (ONOO<sup>-</sup>), nitrogen dioxide (NO<sub>2</sub>) or various derivatives of nitrogen oxide (NO) [6]. FR biosynthesis involve multiple reactions that produce various species in various conditions. Oxidative chain reaction is activated once the super-



Figure 1. Oxidative stress during brain ischemia and traumatic brain injury. TBI, traumatic brain injury; FR, free radicals; OS, oxidative stress

oxide ion is generated. Superoxide anions are the result of mitochondrial leakage, xanthine oxidase activity and arachidonic acid metabolism. The action of superoxide dismutase (SOD) on superoxide anions form hydrogen peroxide and oxygen. HO- is one of the most aggressive FRs. It is synthesised following the reaction between hydrogen peroxide and bivalent iron ions (Fenton Reaction) [7]. By combining the reaction of superoxide anion and nitrogen oxide derivatives, peroxynitrite radicals are produced. The reactivity is increased due to the multitude of reactions that take place in the human body. By protonation of the peroxynitrite radical, peroxynitrous acid is obtained which through denaturation produces significant amounts of hydroxyl radicals and nitric oxide radicals. FR's aggressive reaction on lipids leads to severe physiological imbalances mainly by destroying cellular membrane integrity. Denaturation of lipids occurs by reaction of FR with polyunsaturated fatty acids such as arachidonic acid, linoleic acid, eicosapentaenoic acid, and docosahexaenoic acid. Denaturation reactions follow three important steps, initiation, propagation and termination. In lipid oxidation reactions, initially lipidic radical is being produced (L-). Through its reaction with oxygen, a lipid peroxyl radical is formed (LOO-) which, by attaching to a hydrogen ion (L-H), leads to lipid hydroperoxide (LOOH), a new lipid radical. Following the redox reactions of lipid denaturation, two neurotoxic aldehydes are produced: 4-hydroxynonenal, 2 - propenal. Neurotoxicity is a concequence of their interaction with a number of amino acids: lysine, histidine or cysteine, resulting in inactivation of enzymatic and structural functions of proteins. In redox reactions that involve FR, proteins are also attacked, forming active protein carbonyl species or disulfide bonds.

Radicals formed from this chain of redox reactions attack both the structure of DNA and RNA, seriously affecting the functions of transcription, replication and mRNA translation. Impairment of these functions has direct implications in the decreased ability of the cell survival under stress [8,9].

FRs are responsible for inducing secondary injuries leading to progressive deterioration of the clinical status of patients due to mitochondrial dysfunction, which is responsible for the biosynthesis of peroxynitrite radical, Ca<sup>2+</sup> buffering impariment, pump Na<sup>+</sup> / K<sup>+</sup> - ATP imbalance and accumulation of intracellular calcium, and microvascular systems damage.

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# SPECIFIC BIOMARKERS FOR OXIDATIVE STRESS AND TRAUMATIC BRAIN INJURY

To highlight the pathophysiological effects induced by activation of OS in patients with TBI, a number of specific biomarkers have been studied (Table 1). One of the most studied biomarker of TBI is \$100-calcium binding protein beta (\$100B) [10-12].

Falcone et al, studied the correlation of serum and urine levels of S100B and TBI [13], highlighting the correlation between TBI and increased serum S100B.

A number of specialized studies report a direct and statistically significant correlation between increased serum levels of S100B and several clinical problems such as the magnitude of injury, degree of survival or post-traumatic neurological sequelae. Serum levels of S100B more than 1.14 ng/mL were correlated with an increase in mortality and morbidity [6,11,14,15].

Another biomarker commonly used for identification and scoring the degree of TBI is glial fibrillary acidic protein (GFAP). This is an insoluble protein and is rapidly induced following brain injuries. Hol et al demonstrated the correlation between serum levels of GFAP and different stages of brain injury [16]. Numerous other studies demonstrate that increased serum levels of GFAP are correlated with an increased intracranial pressure (ICP) and decreased cerebral perfusion pressure (CPP) [17-19]. Values greater than 1.5 ng/mL are correlated directly and significantly with decreased survival rate in critical patients with TBI [15,20,21].

Neuron-specific Enola (NSE) is another specific biomarker for TBI [22,23]. Reports show a correlation between serum levels greater than 12.5 ng/mL of NSE and severe brain injury. NSE is considered one of the most specific biomarkers for TBI, its serum levels being correlated with the intensity of TBI. El-Maraghi et al, reported on the importance of NSE in diagnosing TBI and its specificity for brain damage [24]. Following injury, a specific protein called myelin basic protein is released (MBP), and this is used to predict TBI [25]. Studies report that an increase in serum levels of more than 0.3 ng / mL is significantly correlated with an increase in morbidity or mortality for patients with TBI [20].

Products of metabolism of neurotransmitters are also considered important indicators in TBI. Barco et al highlight increased levels of homovanillic acid

Antioxidant	Mechanism of action	References
Vitamin C	reduced neutrophil oxidative burst; endothelial protective effect; de- creases ROS production; prevents vascular leakage; prevents microvas- cular thrombosis; attenuates sepsis, decreases serum concentration of malondialdehyde;	[34, 45, 51, 66, 70]
Vitamin E	inhibits the multiplication of free radicals in lipid peroxidation reac- tions; reduces side effects of hyperhomocysteinemia;	[11, 18, 34, 45, 65]
Vitamin B1	inhibits the multiplication of free radicals;	[12, 23, 34, 56, 56, 56]
N-acetylcysteine	inhibits lipid peroxidation; Anti-inflammatory;	[12, 23, 34, 23, 59]
Resveratrol	Anti-inflammatory; antiplatelet; neuroprotective effects;	[5, 8, 9]
Melatonin	modulation of mitochondrial activity; protection of neuronal mito- chondrial membrane; maintaining normal parameters and activity of Na + / K +-ATP pump; reduced synthesis of free radicals in the mito- chondria;	[3, 4, 12, 11]
Selenium	reduce production of free radicals; inhibit oxidative chain initiation;	[34, 45, 69]
Zinc	reduce production of free radicals;	[12]
U-83836E (second gen- eration lozaroid)	inhibit lipid peroxidation in oxidative chain initiation; minimizes the production of free radicals;	[12, 34, 48, 67]
PEG-SOD (stable PEG- conjugated superoxide dismutase)	improves antioxidant activity of superoxide dismutase enzyme; inhibit the production of free radicals; restores the balance of antioxidant- oxidant;	[12, 34, 56, 65]
methylprednisolone	inhibit lipid peroxidation; mainly used in TBI associated with spinal cord injuries;	[17, 23, 56]

#### Table 1. Antioxidant therapy in traumatic brain injury

(produced by metabolism of dopamine) in patients with TBI [26]. The product of metabolism of serotonin (5-hydroxy-indoleacetic acid) is another biomarker used in assessing the degree of brain injury [20,27,28].

Useful markers in the evaluation and optimization of intensive therapy in TBI are represented by inflammatory markers. The following studies show significant serum levels of pro- and anti- inflammatory cytokines and chemokines. Interleukin 1 (IL-1), interleukin 6 (IL-6), tumour necrosis factor alpha (TNF-alpha) are the most representative in case of TBI [20,29,30]. IL-1 has been extensively studied due to its increased specificity. A number of specialized studies correlate the increased serum levels of IL-1 with poor outcomes [20].

Moreover, studies conducted both on animals and in humans reported a direct and statistically significant correlation between increased levels of IL-6 and the extent of brain damage [12,15,20]. Following secondary injuries induced by OS, a number of specific markers are released. Classification of the severity of OS can be done both directly and indirectly by dosing specific markers.

4-hydroxynonenal [31] is a specific biomarker for redox reactions of lipid denaturation while 8-hydroxy-2-deoxyguanosine is used to assess injuries caused by the effect of FR on DNA and RNA and the resultant expression of DNA oxidative damage [20]. Another biomarker extensively used for detection of OS in the brain is 8-epi-PGF2a [32]. Aggression brought by NO derived free radicals are highlighted with biomarker 3NT. After prolonged action of OS, metallopreoteinases (MMPs) are activated, and result in an increased permeability of the blood-brain barrier.

Numerous studies report that increased activity of MMPs leads to destruction of brain microvascular system. Hohl et al, dosed plasma levels of thiobarbituric acid reactive species (TBARS) and carbonyl groups [33]. TBARS is used as an indirect biochemical markers of lipid peroxidation. Oxidative denaturation of proteins was assessed by analytical determination of plasma carbonyl groups. Hohl et al showed a significant difference in plasma TBARS and carbonyl groups in the first 70 hours after severe TBI [33].

# CIRCULATING MICRORNAS AS INDICATOR FOR OS

MicroRNAs are 19-23 nucleotide-containing biomolecules, representing a small fraction of the total mass of ribonucleic acid (RNA) [34], a number of which can be linked to a series of physiological dysfunctions. Thus, microRNAs extracted from biological samples can be used as a biomarker for diagnosis of physiological and biochemical imbalances, including cancer, neurodegenerative disorders, inflammation, infection with different pathogens, metabolism disorders, oxidative stress and poisoning [35,36]. In TBI, some microR-NAs were identified and used as biomarkers for both primary and secondary injuries induced by OS.

Guroang et al showed the presence of a considerable number of microRNAs species in the case of FR assault on biological systems, including: microRNA-15a microRNA-15b; microRNA-16 microRNA-20a, micro-RNA20b, microRNA-92, microRNA-106, microRNA-139, microRNA-146b, microRNA-155. They concluded that the microRNA-106b and microRNA-20b family was specific for oxidative stress [37]. Suh et al, reported that microRNA-133 is a biomarker specific for neuronal apoptosis induced by OS [38].

# NATURAL ANTIOXIDANT SYSTEMS

The body's natural antioxidant system is represented by a series of enzymes, highly active in inhibiting FR. One of the most effective antioxidant is the superoxide dismutases enzyme systems (SODs) [39]. Depending on the location but also the type of metal ion present at the active site, three types of enzymes have been identified. These are SOD1 (CuZnSOD) identified in the nucleus and cytosol, SOD2 (MnSOD) identified in the mitochondrial matrix and SOD3 (EcSOD) localized extracellulary [40,41]. Antioxidant activity is represented by inhibition of oxidizing activity of superoxide ion and its transformation into molecules of oxygen and hydrogen peroxide [39].

Catalase is another enzyme able to reduce the oxidative effect of hydrogen peroxide by decomposing it to molecules of oxygen and water. Antioxidant activity is due the four porphyrin haem groups present in catalase structure, making it possible to interact with hydrogen peroxide [42,43].

Bio-production of glutathione (GSH) results from the interaction of glutamate and cysteine. The antioxidant activity of GSH is dependent on reduced NADPH as the electron donor glucose-6-phosphate dehydrogenase. GSH antioxidant action maintains redox balance within the normal range [44]. Glutathione peroxidase (GPX), a protein that contains selenium, is responsible for reducing peroxidases. Numerous studies have

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shown a intense decrease in the level of GPX with a concomitant increase in lipid peroxidation in TBI [45]. Glutaredoxins (Grxs) are present in the human organism in two forms. Grx1 is present in the cytosol and intermembranous space and Grx2 is present in the mitochondrial matrix. Both enzymes are responsible for protein deglutathionylation [45]. Another endogenous antioxidant system responsible for breakdown of hydrogen peroxide is the peroxiredoxins (Prxs) [46]. They are ubiquitous thiol peroxidases, identified in six structural forms. Prx1, Prx2 and Prx6 are found in the cytoplasm, Prx4 in the endoplasmic reticulum and Prx3 and Prx5 in mitochondria [47]. An intensely studied antioxidant is heme-oxygenase (HO) [48]. HO is a microsomal enzyme with an important role in ensuring cell cycle of Fe<sup>2+</sup> ions [49]. In addition to changes at the biochemical level, many optical microscopy studies using routine immunohistochemistry stains show a close correlation between the OS and structural changes of the brain and cerebellum.

During haemorrhage, tissue accumulates haem which activates heme oxygenase (HO-1) involved in the metabolism of heme and its conversion to carbon monoxide, iron ions and biliverdin [50]. Biliverdin is a powerful antioxidant and therefore HO-1 expression is considered to be protective against oxidative stress. In subarachnoid haemorrhage, ischemic, traumatic brain injury and neurodegenerative diseases, the presence of HO-1 was demonstrated in microglia, astrocyte and neurons [50,51].

In humans, the presence of microglia, positive for HO-1 was demonstrated six hours after traumatic brain injury [52]. Histopathological changes in the brain and cerebellum depend on the aetiological agent of the primary injury.

Bhalla et al showed differences between the cerebellum and brain histology in rats with brain injuries induced by administration of aluminium compared to control group [53], with disruption of the layers, presence of vacuolar spaces and loss of Purkinje neuron layer. It was noted that lithium treatment partially restored the organ architecture and led to improvement of enzyme activity in the cerebellum in proportion to the administered dose. The authors concluded that lithium played a neuroprotective role [53].

# ANTIOXIDANT THERAPY STRATEGY

Major imbalances between oxidants and antioxidants are associated with prolonged hospital stay, with sig-

nificant contributions to increased mortality and morbidity in critically ill patients. Antioxidant therapy in patients with TBI should address the inhibition of the synthesis of ROS and RNS, blocking of the initiation of lipid peroxidation and inhibition of the propagation of the chemical biochemical [1].

A study by Blass et al showed a significant decrease of plasma micro-nutrients and significantly decreased levels of ascorbic acid, retinol, 25-hydroxycholecalciferos, beta-carotene, zinc and selenium in patients with multiple trauma. It further revealed significant differences between antioxidant capacity between the control group and the group with trauma [54].

One of the most studied compounds having an antioxidative effect for neuronal cells is N-acetyl-5-methoxytryptamine (melatonin) [55]. Under normal physiological conditions endogenous production of melatonin is stimulated by a number of precursors including serotonin, arylalkylamine N – acetyltransferase and hydroxyl tryptophan [3]. Together with specific energy and metabolic imbalances, melatonin production is affected, resulting in significant decreases in its serum levels. The antioxidant activity of melatonin is modulated by specific receptors (MT1 and MT2) present on the surface of the neuron [3,56].

It is well documented that by interacting with the mitochondrial membrane and by optimizing oxidative phosphorylation and maintaining normal levels of ATP, melatonin is responsible for protecting brain membranes and mitocondrial activity. Several studies show that melatonin administration considerably decreases blood-brain barrier permeability (BBB), resulting in decreased cerebral oedema. It was also shown that high doses of melatonin increase the activity of endogenous antioxidant enzymes [3,57].

Another antioxidant used mainly in head trauma associated with spinal cord injuries is methylprednisolone. This glucocorticoid steroid is recommended in TBI associated with spinal cord injuries due to inhibition of lipid peroxidation. Studies suggest the administration of 30 mg / kg as a bolus (intravenously) followed by infusion during 23 hours of 5.4 mg/kg [58].

U-83836 is a pharmaceutical compound described as second generation lazaroid and its effects have been reported in many studies. Antioxidant effects of this preparation are due to inhibition of lipid peroxyl radical (LOO-) which blocks the redox chain reaction [59].

Superoxide ion is one of the most aggressive FR. Its neurotoxicity is given by a number of factors, including

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biosynthesis accelerated by neutrophils, increased production during the inflammation response, increased reactivity of nitric oxide and peroxynitrite production. SOD is the only enzyme with antioxidant capacity able to inhibit oxidative activity of superoxide ion by converting it into oxygen and hydrogen peroxide molecules [39].

Numerous studies on the incorporation of SOD enzyme in various matrices able of controlling the release of active substances, reported significant decreases in biochemical markers specific to OS. Porfire et al, demonstrated the antioxidant of SOD entrapped in liposomal formulation with polyethylene glycol matrix [60].

A substance with high antioxidant capacity extensively studied lately is resveratrol (3,4,5 - trihydroxytrans-stilbene) [61]. This is a polyphenol compound found mainly in black grapes. The antioxidant activity of resveratrol is mainly associated with the modulation of mitochondrial activity [62].

Song et al studied the antioxidant effects of resveratrol, highlighting that its main feature is to regulate the expression of sirtuin 1 (NAD - dependent histone deacetylase class III), responsible for the modulation of metabolism of mitochondria, and PGC - 1alpha (proliferation activated receptor coactivator - 1alpha), responsible for mithocondrial biogenesis [63]. In several studies, due to its neuroprotective properties, resveratrol is recommended for use to combat the side effects induced by the OS.

The beneficial effects of antioxidant therapy with vitamin C were observed in a number of studies on ischemia reperfusion syndromes. High doses of vitamin C have beneficial effects, especially on the lung, liver and brain [64]. Intravenous administration of high doses lead to reduced injuries caused by the OS with reduced neutrophil oxidative burst [65] and endothelial protective effects [66]. There is a decreased biosynthesis of free radicals, protective mitochondrial activity, minimization of energy failure, attenuation of sepsis, reduction of microvascular thrombosis rate, normalization of coagulation and prevention of vascular leakage [67].

Nathens et al studying the antioxidant therapy in the critically ill, reported a number of benefits following the association of vitamin C with Vitamin E [68]. The intravenous administration of 1000 mg/day of vitamin C and 1000 IU Vitamin E given orally produced significant decrease in the duration of mechanical ventilation, ICU length of stay, multiple organ failure and

mortality [68]. Also, the combination of vitamin E with selenium resulted in reducing OS due to synergic antioxidative activity. Several studies indicate that the administration of selenium with vitamin E inhibits lipid peroxidation.

Administration of folate, vitamin B12, B6 and B1 have antioxidant effects due to normalizing the plasma level of homocysteine, thus lowering the incidence of cerebral venous thrombosis [69].

Şenol et al, demonstrated that administration of Nacetylcysteine together with selenium lead to normal serum values of OS biomarkers [70]. Furthermore, Nacetylcysteine is responsible for regulation of erythrocyte glutathione peroidase, minimizing the excitotoxic agents activity and inhibition of redox chain of lipid peroxidation [70].

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Following TBI, secondary injuries due to generated changes, contribute significantly to he survival rate of critical ill patients.

OS and neurodenaturation of specific factors often lead to death.

Research on FRs and their role in secondary injuries aims to maximize, optimise and diversify antioxidant strategies. In conclusion, the implementation of targeted antioxidant therapy in critically ill trauma patients with severe head injury is recommended due to a reduction in the number of post-traumatic complications. However, further studies are required to arrive at a definitive and appropriate antioxidant therapy.

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- Bains M, Hall ED. Antioxidant therapies in traumatic brain and spinal cord injury. Biochim Biophys Acta - Mol Basis Dis. 2012;1822:675–84.
- 2. Humberto J, Mantilla M, Fernando L, Arboleda G. Revista Colombiana de Anestesiología Anesthesia for patients with traumatic brain injury. Colomb J Anesthesiol. 2015;43:3–8.
- Ashafaq M, Varshney L, Khan MHA, et al. Neuromodulatory effects of hesperidin in mitigating oxidative stress in streptozotocin induced diabetes. Biomed Res Int. 2014;2014: 249031.

#### Available online at: www.jccm.ro

- Sies H. Redox Biology Oxidative stress: a concept in redox biology and medicine. Redox Biol. 2015;4:180–3.
- Ribeiro A, Ferraz-de-Paula V, Pinheiro ML, et al. Cannabidiol, a non-psychotropic plant-derived cannabinoid, decreases inflammation in a murine model of acute lung injury: Role for the adenosine A2A receptor. Eur J Pharmacol. 2012;678:78– 85.
- Abdul-Muneer PM, Chandra N, Haorah J. Interactions of Oxidative Stress and Neurovascular Inflammation in the Pathogenesis of Traumatic Brain Injury. Mol Neurobiol. 2015;51:966-79
- Bibi H, Vinokur V, Waisman D, et al. Zn/Ga-DFO iron-chelating complex attenuates the inflammatory process in a mouse model of asthma. Redox Biol. 2014;2:814–9.
- 8. Hall ED, Vaishnav R a., Mustafa AG. Antioxidant Therapies for Traumatic Brain Injury. Neurotherapeutics. 2010;7:51–61.
- Miller DM, Singh IN, Wang JA, Hall ED. Nrf2 ARE activator carnosic acid decreases mitochondrial dysfunction, oxidative damage and neuronal cytoskeletal degradation following traumatic brain injury in mice. Exp Neurol. 2015;264:103–10.
- Rodríguez-Rodríguez A, Egea-Guerrero JJ, León-Justel A, et al. Role of S100B protein in urine and serum as an early predictor of mortality after severe traumatic brain injury in adults. Clin Chim Acta. 2012;414:228–33.
- Strathmann FG, Schulte S, Goerl K, Petron DJ. Blood-based biomarkers for traumatic brain injury: evaluation of research approaches, available methods and potential utility from the clinician and clinical laboratory perspectives. Clin Biochem. 2014;47:876–88.
- Kumar RG, Diamond ML, Boles JA, et al. Acute CSF interleukin-6 trajectories after TBI: Associations with neuroinflammation, polytrauma, and outcome. Brain Behav Immun. 2014;45:253– 62.
- Falcone T, Janigro D, Lovell R, et al. S100B blood levels and childhood trauma in adolescent inpatients. J Psychiatr Res. 2014;62:14–22.
- Cervellin G, Benatti M, Carbucicchio A, et al. Serum levels of protein S100B predict intracranial lesions in mild head injury. Clin Biochem. 2012;45:408–11.
- Dal-Pizzol F, Ritter C, Cassol-Jr OJ, et al. Oxidative mechanisms of brain dysfunction during sepsis. Neurochem Res. 2010;35:1–12.
- Hol EM, Pekny M. Glial fibrillary acidic protein (GFAP) and the astrocyte intermediate filament system in diseases of the central nervous system. Curr Opin Cell Biol. 2015;32C:121–30.
- 17. Feneberg E, Steinacker P, Lehnert S, Böhm B, Mayer G, Otto M. Elevated glial fibrillary acidic protein levels in the cerebrospinal fluid of patients with narcolepsy. Sleep Med. 2013;14:692–4.
- Tennakoon AH, Izawa T, Wijesundera KK, et al. Immunohistochemical characterization of glial fibrillary acidic protein (GFAP)-expressing cells in a rat liver cirrhosis model induced by repeated injections of thioacetamide (TAA). Exp Toxicol Pathol. 2015;67:53–63.

#### The Journal of Critical Care Medicine 2015;1(3) $\bullet$ 89

- Wang H, Zhang P, Chen W, Feng D, Jia Y, Xie L. Serum microRNA signatures identified by Solexa sequencing predict sepsis patients' mortality: A prospective observational study. PLoS One. 2012;7:1–9.
- Abdul-Muneer PM, Schuetz H, Wang F, et al. Induction of oxidative and nitrosative damage leads to cerebrovascular inflammation in an animal model of mild traumatic brain injury induced by primary blast. Free Radic Biol Med. 2013;60:282– 91.
- Petzold A. Glial fibrillary acidic protein is a body fluid biomarker for glial pathology in human disease. Brain Res. 2014;1600:17– 31.
- Kärkelä J, Bock E, Kaukinen S. CSF and serum brain-specific creatine kinase isoenzyme (CK-BB), neuron-specific enolase (NSE) and neural cell adhesion molecule (NCAM) as prognostic markers for hypoxic brain injury after cardiac arrest in man. J Neurol Sci. 1993;116:100–9.
- 23. Prasad KN, Bondy SC. Common biochemical defects linkage between post-traumatic stress disorders , mild traumatic brain injury (TBI) and penetrating TBI. Brain Res. 2015;1599:103–14.
- 24. El-Maraghi S, Yehia H, Hossam H, Yehia A, Mowafy H. The prognostic value of neuron specific enolase in head injury. Egypt J Crit Care Med. 2013;1:25–32.
- 25. Klevay LM. Myelin and traumatic brain injury: the copper deficiency hypothesis. Med Hypotheses. 2013;81:995–8.
- 26. Barco S, Gennai I, Reggiardo G, et al. Urinary homovanillic and vanillylmandelic acid in the diagnosis of neuroblastoma: report from the Italian Cooperative Group for Neuroblastoma. Clin Biochem. 2014;47:848–52.
- Dash PK, Zhao J, Hergenroeder G, Moore AN. Biomarkers for the diagnosis, prognosis, and evaluation of treatment efficacy for traumatic brain injury. Neurotherapeutics. 2010;7:100–14.
- LeWitt P, Schultz L, Auinger P, Lu M. CSF xanthine, homovanillic acid, and their ratio as biomarkers of Parkinson's disease. Brain Res. 2011;1408:88–97.
- 29. Homsi S, Federico F, Croci N, et al. Minocycline effects on cerebral edema: Relations with inflammatory and oxidative stress markers following traumatic brain injury in mice. Brain Res. 2009;1291:122–32.
- Zampieri FG, Kellum J A, Park M, et al. Relationship between acid-base status and inflammation in the critically ill. Crit Care. 2014;18:R154.
- 31. Ansari M a., Roberts KN, Scheff SW. Oxidative stress and modification of synaptic proteins in hippocampus after traumatic brain injury. Free Radic Biol Med. 2008;45:443–52.
- 32. Wannhoff A, Bölck B, Kübler AC, Bloch W, Reuther T. Oxidative and nitrosative stress and apoptosis in oral mucosa cells after ex vivo exposure to lead and benzo[a]pyrene. Toxicol In Vitro. 2013;27:915–21.
- Hohl A, Gullo JDS, Silva CCP, et al. Plasma levels of oxidative stress biomarkers and hospital mortality in severe head injury: A multivariate analysis. J Crit Care. 2012;27:523.e11–523.e19.
- 34. Ji H-H, Huang G-L, Yin H-X, Xu P, Luo S-Y, Song J-K. Association

#### 90 • The Journal of Critical Care Medicine 2015;1(3)

between microRNA-196a2 rs11614913, microRNA-146a rs2910164, and microRNA-423 rs6505162 polymorphisms and esophageal cancer risk: A meta-analysis. Meta Gene. 2015;3:14–25.

- Muraoka T, Soh J, Toyooka S, et al. The degree of microRNA-34b/c methylation in serum-circulating DNA is associated with malignant pleural mesothelioma. Lung Cancer. 2013;82:485– 90.
- Li Y, Dalli J, Chiang N, Baron RM, Quintana C, Serhan CN. Plasticity of leukocytic exudates in resolving acute inflammation is regulated by MicroRNA and proresolving mediators. Immunity. 2013;39:885–98.
- Li G, Luna C, Qiu J, Epstein DL, Gonzalez P. Alterations in microRNA expression in stress-induced cellular senescence. Mech Ageing Dev. 2009;130:731–41.
- Suh JH, Choi E, Cha M-J, et al. Up-regulation of miR-26a promotes apoptosis of hypoxic rat neonatal cardiomyocytes by repressing GSK-3β protein expression. Biochem Biophys Res Commun. 2012;423:404–10.
- 39. Miller A-F. Superoxide dismutases: ancient enzymes and new insights. FEBS Lett. 2012;586:585–95.
- Gerbaud P, Petzold L, Thérond P, Anderson WB, Evain-Brion D, Raynaud F. Differential regulation of Cu, Zn- and Mnsuperoxide dismutases by retinoic acid in normal and psoriatic human fibroblasts. J Autoimmun. 2005;24:69–78.
- Pilon M, Ravet K, Tapken W. The biogenesis and physiological function of chloroplast superoxide dismutases. Biochim Biophys Acta. 2011;1807:989–98.
- Comar JF, Babeto De Sá-Nakanishi A, De Oliveira AL, et al. Oxidative state of the liver of rats with adjuvant-induced arthritis. Free Radic Biol Med. 2013;58:144–53.
- Rahman I, Adcock IM. Oxidative stress and redox regulation of lung inflammation in COPD. Eur Respir J. 2006;28:219–42.
- 44. Lazzarino G, Di Pietro V, Lazzarino G, et al. Neuroglobin expression and oxidant/antioxidant balance after graded traumatic brain injury in the rat. Free Radic Biol Med. 2014;69:258–64.
- 45. Hybertson BM, Gao B, Bose SK, McCord JM. Oxidative stress in health and disease: The therapeutic potential of Nrf2 activation. Mol Aspects Med. 2011;32:234–46.
- Elkharaz J, Ugun-Klusek A, Constantin-Teodosiu D, et al. Implications for oxidative stress and astrocytes following 26S proteasomal depletion in mouse forebrain neurones. Biochim Biophys Acta- Mol Basis Dis. 2013;1832:1959–68.
- Li L, Zhu K, Liu Y, et al. Targeting Thioredoxin-1 With Sirna Exacerbates Oxidative Stress Injury After Cerebral Ischemia / Reperfusion in Rats. 2015;284:815–23.
- Andrés NC, Fermento ME, Gandini NA, et al. Heme oxygenase-1 has antitumoral effects in colorectal cancer: involvement of p53. Exp Mol Pathol. 2014;97:321–31.
- Yu JH, Cho SO, Lim JW, Kim N, Kim H. Ataxia telangiectasia mutated inhibits oxidative stress-induced apoptosis by

regulating heme oxygenase-1 expression. Int J Biochem Cell Biol. 2015;60:147–56.

- 50. Namba F, Go H, Murphy J A., et al. Expression level and subcellular localization of heme oxygenase-1 modulates its cytoprotective properties in response to lung injury: A mouse model. PLoS One. 2014;9:1–11.
- 51. Kurtz P, Claassen J, Helbok R, et al. Systemic glucose variability predicts cerebral metabolic distress and mortality after subarachnoid hemorrhage: a retrospective observational study. Crit Care. 2014;18:R89.
- Beschorner R, Adjodah D, Schwab JM, et al. Long-term expression of heme oxygenase-1 (HO-1, HSP-32) following focal cerebral infarctions and traumatic brain injury in humans. Acta Neuropathol. 2000;100:377–84.
- 53. Bhalla P, Dhawan DK. Protective Role of Lithium in Ameliorating the Aluminium-induced Oxidative Stress and Histological Changes in Rat Brain. Cell Mol Neurobiol. 2009;29:513–21.
- 54. Blass SC, Goost H, Tolba RH, et al. Time to wound closure in trauma patients with disorders in wound healing is shortened by supplements containing antioxidant micronutrients and glutamine: A PRCT. Clin Nutr. 2012;31:469–75.
- 55. Şener G, Toklu H, Kapucu C, et al. Melatonin protects against oxidative organ injury in a rat model of sepsis. Surg Today. 2005;35:52–9.
- Dehghan F, Khaksari Hadad M, Asadikram G, Najafipour H, Shahrokhi N. Effect of melatonin on intracranial pressure and brain edema following traumatic brain injury: Role of oxidative stresses. Arch Med Res. 2013;44:251–8.
- 57. Yürüker V, Naz M, Nilgün Ş. Reduction in traumatic brain injuryinduced oxidative stress, apoptosis, and calcium entry in rat hippocampus by melatonin : Possible involvement of TRPM2 channels. Metab Brain Dis. 2015;30:223–31.
- Bhalla a., Singhal M, Suri V, Malhotra S, Shafiq N, Varma S. Methylprednisolone in dengue patients with alarm signs: The MIDWAS study. Int J Infect Dis. 2014;21:323.
- 59. Grasbon-Frodl EM, Nakao N, Brundin P. The lazaroid U-83836E improves the survival of rat embryonic mesencephalic tissue stored at 4°C and subsequently used for cultures or intracerebral transplantation. Brain Res Bull. 1996;39:341–7.
- 60. Porfire AS, Leucuţa SE, Kiss B, Loghin F, Pârvu AE. Investigation into the role of Cu/Zn-SOD delivery system on its antioxidant and antiinflammatory activity in rat model of peritonitis. Pharmacol Reports. 2014;66:670–6.
- Schmatz R, Perreira LB, Stefanello N, et al. Effects of resveratrol on biomarkers of oxidative stress and on the activity of delta aminolevulinic acid dehydratase in liver and kidney of streptozotocin-induced diabetic rats. Biochimie. 2012;94:374– 83.
- 62. Koz ST, Etem EO, Baydas G, et al. Effects of resveratrol on blood homocysteine level, on homocysteine induced oxidative stress, apoptosis and cognitive dysfunctions in rats. Brain Res. 2012;1484:29–38.
- 63. Song L, Chen L, Zhang X, Li J, Le W. Resveratrol Ameliorates

#### Available online at: www.jccm.ro

Motor Neuron Degeneration and Improves Survival in SOD1 G93A Mouse Model of Amyotrophic Lateral Sclerosis. Biomed Res Int. 2014;2014:483501.

- 64. Gao J, Koshio S, Ishikawa M, Yokoyama S, Mamauag REP. Interactive effects of vitamin C and E supplementation on growth performance, fatty acid composition and reduction of oxidative stress in juvenile Japanese flounder Paralichthys olivaceus fed dietary oxidized fish oil. Aquaculture. 2014;422-423:84–90.
- 65. Fox ED, Heffernan DS, Cioffi WG, Reichner JS. Neutrophils from critically ill septic patients mediate profound loss of endothelial barrier integrity. Crit Care. 2013;17:R226.
- Chen Q, Jones D, Stone P, Ching LM, Chamley L. Vitamin C Enhances Phagocytosis of Necrotic Trophoblasts by Endothelial Cells and Protects the Phagocytosing Endothelial Cells from Activation. Placenta. 2009;30:163–8.

#### The Journal of Critical Care Medicine 2015;1(3) • 91

- 67. Oudemans-van Straaten HM, Man A, de Waard MC. Vitamin C revisited. Crit Care. 2014;18:460.
- 68. Nathens AB, Neff MJ, Jurkovich GJ, et al. Randomized, prospective trial of antioxidant supplementation in critically ill surgical patients. Ann Surg. 2002;236:814–22.
- 69. Nagaraja D, Noone ML, Bharatkumar VP, Christopher R. Homocysteine, folate and vitamin B12 in puerperal cerebral venous thrombosis. J Neurol Sci. 2008;272:43–7.
- Şenol N, NazIroğlu M, Yürüker V. N-acetylcysteine and selenium modulate oxidative stress, antioxidant vitamin and cytokine values in traumatic brain injury-induced rats. Neurochem Res. 2014;39:685–92.
- Navarro-Yepes J, Zavala-Flores L, Anandhan A, et al. Antioxidant gene therapy against neuronal cell death. Pharmacol Ther. 2014;142:206–30.